

# A multidomain hub anchors the chromosome segregation and chemotactic machinery to the bacterial pole

Yoshiharu Yamaichi,<sup>1,2,5</sup> Raphael Bruckner,<sup>1,2,5,6</sup> Simon Ringgaard,<sup>1,2</sup> Andrea Möll,<sup>1,2</sup> D. Ewen Cameron,<sup>2,7</sup> Ariane Briegel,<sup>3</sup> Grant J. Jensen,<sup>3,4</sup> Brigid M. Davis,<sup>1,2</sup> and Matthew K. Waldor<sup>1,2,4,8</sup>

<sup>1</sup>Division of Infectious Diseases, Brigham and Women's Hospital, <sup>2</sup>Department of Microbiology and Immunobiology, Harvard Medical School, Boston, Massachusetts 02115, USA; <sup>3</sup>Division of Biology, California Institute of Technology, Pasadena, California 91125, USA; <sup>4</sup>Howard Hughes Medical Institute, Boston, Massachusetts 02115, USA

The cell poles constitute key subcellular domains that are often critical for motility, chemotaxis, and chromosome segregation in rod-shaped bacteria. However, in nearly all rods, the processes that underlie the formation, recognition, and perpetuation of the polar domains are largely unknown. Here, in *Vibrio cholerae*, we identified HubP (hub of the pole), a polar transmembrane protein conserved in all vibrios, that anchors three ParA-like ATPases to the cell poles and, through them, controls polar localization of the chromosome origin, the chemotactic machinery, and the flagellum. In the absence of HubP, *oriCI* is not targeted to the cell poles, chemotaxis is impaired, and a small but increased fraction of cells produces multiple, rather than single, flagella. Distinct cytoplasmic domains within HubP are required for polar targeting of the three ATPases, while a periplasmic portion of HubP is required for its localization. HubP partially relocalizes from the poles to the mid-cell prior to cell division, thereby enabling perpetuation of the polar domain in future daughter cells. Thus, a single polar hub is instrumental for establishing polar identity and organization.

[Keywords: *Vibrio cholerae*; cell polarity; chemotaxis; chromosome segregation; motility]

Supplemental material is available for this article.

Received June 29, 2012; revised version accepted August 27, 2012.

The formation and recognition of subcellular domains is critical for numerous cellular processes, even in relatively simple unicellular organisms such as bacteria (Shapiro et al. 2009; Rudner and Losick 2010; Thanbichler 2011). Without an ability to establish intracellular landmarks, key events in the bacterial cell cycle, such as chromosome segregation and formation of the cell division plane, are instead subject to stochastic variation that can lead to detrimental results. Bacterial morphology and motility also often depend on positional recognition; e.g., for correct localization of pili and flagella. A few determinants of such subcellular patterns have been well studied, such as

MinCDE, which enable a number of bacterial species to form a division plane at the mid-cell (Raskin and de Boer 1997; Lutkenhaus 2007). In Gram-positive bacteria and actinomycetes, DivIVA recruits other proteins to the cell poles and septum and thereby regulates several cell processes (Marston et al. 1998; Ben-Yehuda et al. 2003; Flärdh 2010). Furthermore, in the differentiating  $\alpha$ -proteobacterium *Caulobacter crescentus*, TipN and PopZ are markers and determinants of pole formation (Huitema et al. 2006; Lam et al. 2006; Bowman et al. 2008; Ebersbach et al. 2008). However, in most bacteria, the means by which subcellular domains are generated or recognized remain unknown.

In the Gram-negative rod *Vibrio cholerae*, as in other rod-shaped bacteria, the cell poles are key subcellular domains, and the bacterium has the ability to distinguish poles from sides and the old pole from the new (most recently generated) pole. For example, following cell division, the origin region of the larger of *V. cholerae*'s two chromosomes (*oriC* of *chrI*, *oriCI*) is always found at the old pole, as is an array of chemoreceptors with associated

<sup>5</sup>These authors contributed equally to this work.

Present addresses: <sup>6</sup>Department of Cell Biology, Harvard Medical School, 240 Longwood Avenue, Boston, MA 02115, USA; <sup>7</sup>Department of Biomedical Engineering, 44 Cummings Street, Boston University, Boston, MA 02215, USA.

<sup>8</sup>Corresponding author

E-mail [mwaldor@rics.bwh.harvard.edu](mailto:mwaldor@rics.bwh.harvard.edu)

Article is online at <http://www.genesdev.org/cgi/doi/10.1101/gad.199869.112>.

chemosignaling proteins, and the bacterial flagellum also develops at this site. We found that targeting of *oriCI* to the pole is dependent on ParA1 (Fogel and Waldor 2006), a homolog of bacterial plasmid partitioning proteins, and that the localization of the chemotactic apparatus is also regulated by a ParA homolog, ParC (Ringgaard et al. 2011). Flagellar assembly is regulated by a third ParA-like protein, FlhG (Correa et al. 2005; Kusumoto et al. 2008), raising the possibility that these proteins rely on related processes to govern spatial patterning. Notably, each protein forms a focus at the old pole in newborn cells, although their distribution is not always limited to this site (Fogel and Waldor 2006; Kusumoto et al. 2008; Ringgaard et al. 2011). As the cell cycle progresses and the cell prepares for division, which will yield a “new” old pole in one of the daughter cells, ParA1, ParC, and FlhG all adopt a bipolar pattern. However, the means by which their distribution is initially determined and subsequently shifted has not been established. *V. cholerae* (a  $\gamma$ -proteobacterium) lacks apparent homologs of TipN and PopZ; in fact, these polar determinants are restricted to a subset of  $\alpha$ -proteobacteria (Lam et al. 2006; Bowman et al. 2008; Ebersbach et al. 2008).

ParA family proteins are typically relatively small, cytoplasmic proteins that contain a deviant Walker AAA ATPase motif. They often have the ability to self-associate; e.g., into polymers that may push or pull DNA and thereby promote its equal distribution between daughter cells. ParA family proteins have also been found to govern the localization of the division site (e.g., via MinD and MipZ) (Thanbichler and Shapiro 2006; Lutkenhaus 2007), type IV pili (e.g., via TadZ/CpaE) (Viollier et al. 2002; Perez-Cheeks et al. 2012; Xu et al. 2012), and conjugative transfer machinery (via VirC1) (Atmakuri et al. 2007). They often have a dynamic distribution within the cell, and this distribution is key for proper function. In most cases, the proteins' ability to bind and/or hydrolyze ATP is critical for their subcellular localization and function (Szardenings et al. 2011; Lutkenhaus 2012). It should be noted, however, that not all ParA-like proteins, even in *V. cholerae*, form foci at the cell poles. The small chromosome (chrII)-associated protein ParA2 and MinD are oscillatory and more diffusely distributed throughout much of the cell (Raskin and de Boer 1999; Fogel and Waldor 2006). ParA proteins associated with plasmid segregation have also been observed to oscillate. The distinct localization patterns of these proteins are likely determined by the specific protein partners with which each interacts.

We performed a genetic screen to identify factors that contribute to the establishment of polar identity in *V. cholerae* and found that a single protein, VC0998 (henceforth called HubP, for hub of the pole), is required for the proper polar localization of ParA1, ParC, and FlhG. In the absence of HubP, *oriCI* is not targeted to the cell poles, chemotaxis is impaired due to the absence or mislocalization of polar chemotactic receptor arrays and their associated signaling proteins, and cells have an increased propensity to produce multiple, rather than single, flagella. HubP, like the three ParA family ATPases, is localized to

the cell poles; however, it routinely marks both poles—rather than the single (old) pole typically observed for the other proteins in young cells—and can migrate between the two poles. Despite the homology between ParA1, ParC, and FlhG, their polar targeting by HubP appears to rely on different mechanisms, and a distinct region of HubP is needed to localize each client protein. Thus, HubP is a multifaceted scaffold necessary for the establishment of polar identity and organization; it is a junction point around which multiple polar processes are oriented.

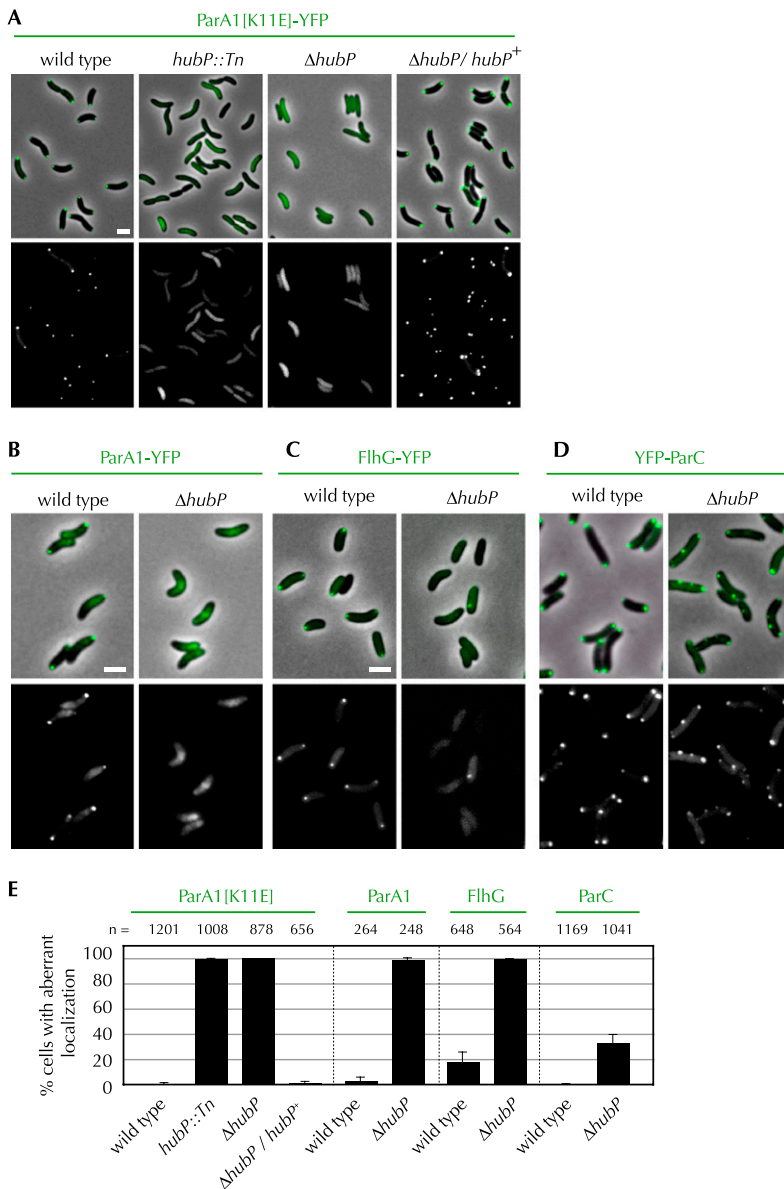
## Results

### *Identification of HubP, an organizer of polar features and processes*

Our screen for pole-organizing factors, which was similar to a screen in *C. crescentus* (Huitema et al. 2006), was premised on the idea that *V. cholerae* lacking a polar landmark protein is likely to have impaired motility/chemotaxis, since its flagella and/or chemotaxis proteins may be mislocalized. Therefore, we focused our analysis on genes that had previously been linked to a motility defect (Cameron et al. 2008) yet did not have an obvious connection to chemotaxis or flagellar assembly. A set of 91 mutants was transformed with an expression construct for a previously described polar marker protein, ParA1[K11E]-YFP, which, in contrast to wild-type ParA1-YFP, forms discrete bipolar foci throughout the cell cycle (Ringgaard et al. 2011). Transformants were individually screened using fluorescence microscopy to identify strains in which this protein is mislocalized.

We found that disruption of a previously undescribed gene, *vc0998* (*hubP*), caused ParA1[K11E] to be diffusely distributed throughout cells, rather than displaying its typical bipolar distribution (Fig. 1A,E). This pattern was observed in both a transposon insertion mutant (*hubP::Tn*) and a strain with an in-frame deletion of *hubP*. Furthermore, wild-type localization of ParA1[K11E] was restored in the  $\Delta$ *hubP* strain by expression of the protein in *trans*, providing strong evidence that polar targeting of ParA1[K11E] is dependent on the presence of HubP.

HubP is also required for the proper cellular positioning of several additional polar factors. In wild-type cells, at least a portion of ParA1, ParC, and FlhG is found within polar foci at either one or both poles, depending on the age of the cell. However, in a strain lacking *hubP*, the distribution of these three proteins was markedly different than in wild-type cells (Fig. 1B–E). ParA1 was diffusely distributed throughout the cell (Fig. 1B), and FlhG was either diffusely distributed or found in a nonpolar focus (Fig. 1C); neither protein formed polar foci. ParC formed polar foci in a subset of *hubP* cells (Fig. 1D); however, ~50% of cells contained nonpolar ParC foci either instead of or in addition to a polar focus. An increase in diffusely distributed ParC was also evident in the *hubP* cells. Thus, HubP appears to be important for the proper subcellular distribution of three ParA-type ATPases in *V. cholerae*. However, it is not required for the correct cellular targeting of all such paralogous proteins. The

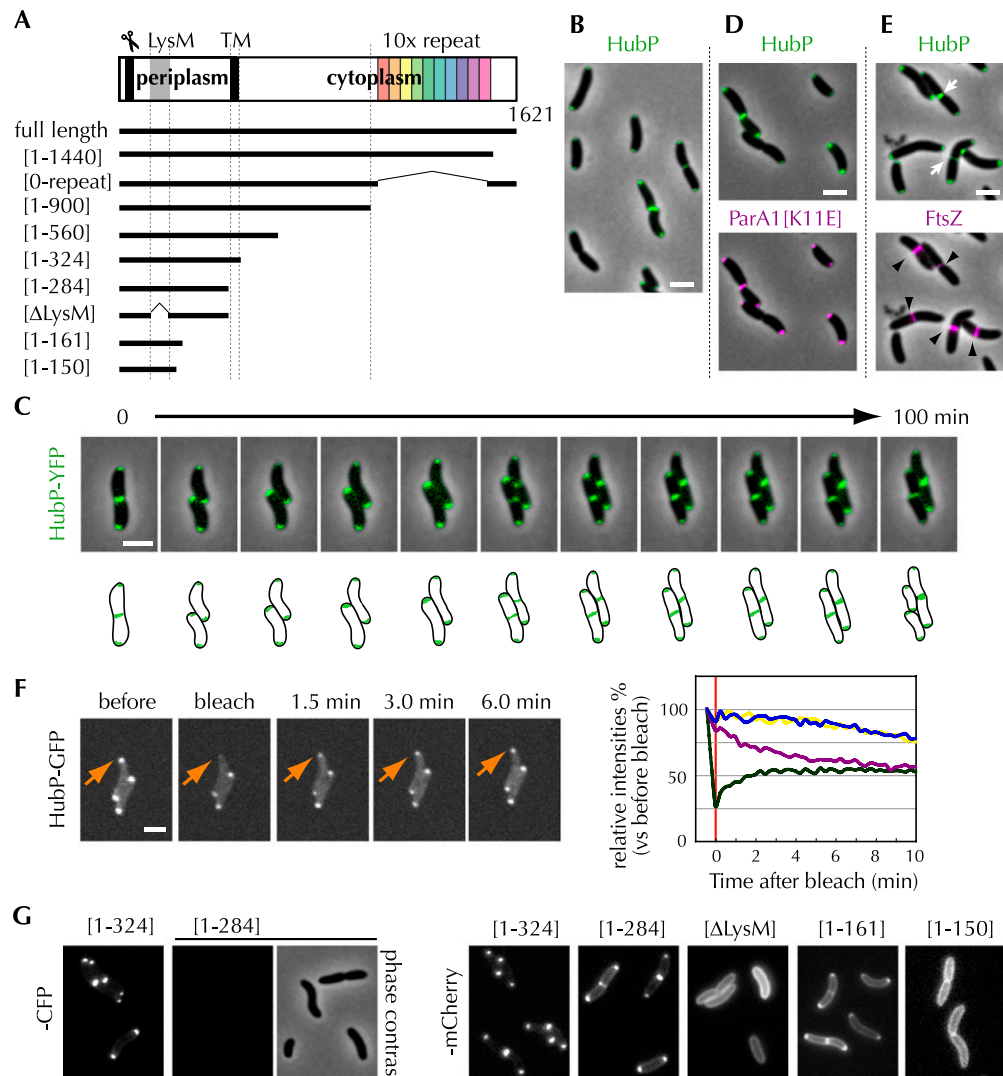


**Figure 1.** HubP is a determinant of polar protein localization. (A–D) Subcellular localization of polar proteins fused to YFP (pseudocolored in green with phase contrast in the top panels) in indicated *V. cholerae* strains. (A) ParA1[K11E]. (B) ParA1. (C) FlhG. (D) ParC. Pictures shown are representative fields. Bar, 2  $\mu$ m. (E) Percentage of cells with mislocalized polar proteins. For ParA1[K11E], ParA1, and FlhG, mislocalization refers to cells lacking a polar focus, and for ParC, it refers to cells that contain a nonpolar focus. Means, standard deviations, and total number of cells counted (*n*) are shown. See also Supplemental Fig. S1.

distribution of ParA2, which mediates chrII partitioning, and of MinD, which likely specifies the *V. cholerae* division site, was not altered in the absence of HubP (Supplemental Fig. S1). Additionally, the absence of HubP did not cause detectable changes in cell shape or cell size and did not impair cell growth.

Sequence and genomic analyses revealed several notable attributes of the HubP protein. HubP is quite large (1621 amino acids, or ~178 kDa) and extremely acidic (pI = 3.22). Its N terminus appears to contain a signal sequence, a potential LysM peptidoglycan (PG)-binding domain, and a single transmembrane domain. Its C terminus, which is predicted to reside within the cytoplasm, contains 10 copies of an imperfect 46-amino-acid repeat, in which 19 of the consensus amino acids are acidic (Fig. 2A; Supplemental Fig. S2). The protein is fairly

well conserved among vibrio and photobacteria species (*Vibrionaceae/Photobacteriaceae*); for example, homologs with >40% identity are present in *Vibrio vulnificus* and *Vibrio parahaemolyticus*, and *Vibrio fischeri* encodes a protein with 27.5% identity. Homologs appear to have similar overall structures, although the sizes and copy numbers of their acidic repeat sequences vary significantly (Supplemental Table S1; Supplemental Fig. S2). Outside of the *Vibrionaceae*, homologs are found principally in the  $\gamma$ -proteobacteria, where they are significantly less conserved, but many appear to be encoded within a similar chromosomal neighborhood. Some of these proteins (e.g., FimV of *Pseudomonas aeruginosa*) have been found to be associated with the assembly of type IV pili and pilus-associated twitching motility (Semmler et al. 2000; Wehbi et al. 2011).



**Figure 2.** HubP localization and its determinants. (A) Schematic of the HubP polypeptide (top) and truncation mutants (bottom) used in this study. (Scissors) Signal sequence; (LysM) LysM domain; (TM) transmembrane domain; (10× repeat) repeat sequence (see the text). (B) Subcellular localization of HubP-CFP in *ΔhubP V. cholerae*. (C, top) Time-lapse images of plasmid-borne HubP-YFP in *ΔhubP V. cholerae*. A schematic representation is drawn below. (D, E) Subcellular localization of HubP-CFP and ParA1[K11E]-YFP (D) and FtsZ-YFP (E) in *ΔhubP V. cholerae*. In E, HubP and FtsZ at mid-cell are indicated with arrowheads and arrows, respectively. (F) FRAP experiment of HubP-GFP. Pictures from a representative experiment (left) are shown, along with a graph of average signal intensities over time, based on analysis of 30 cells (right). The bleached focus is indicated by an arrow in the pictures and by the green line in the graph. The magenta line shows the average intensity of the focus at the pole opposite the bleached pole. Pale blue and yellow lines show the intensities of polar foci from unbleached (control) cells. (G) Fluorescence of -CFP and -mCherry fusions to truncated HubP in *V. cholerae*. Representative fields are shown. Bar, 2 μm.

### HubP is directed to the *V. cholerae* pole and mid-cell by its periplasmic domain

Given the effect of HubP deficiency on subcellular targeting of polar proteins, we assessed whether HubP itself is present at the *V. cholerae* cell poles. For these experiments, we used fluorescence microscopy and visualized either ectopically or chromosomally expressed HubP-CFP (or YFP/GFP). The fusion proteins, which are functional, as they direct ParA1[K11E]-YFP to a polar position in the *hubP* mutant, displayed a bipolar distribution,

even in small (young) cells (Fig. 2B–D). Thus, HubP appears to mark the new pole prior to the arrival of the ParA family proteins, which are typically not bipolar in young cells (Fogel and Waldor 2006; Ringgaard et al. 2011). HubP was also detectable at the mid-cell in a subset of cells, where it always colocalized with FtsZ, suggesting that it arrives at this site as cells prepare to divide (Fig. 2E). Time-lapse analyses confirmed that HubP is not present at the mid-cell in young cells, but instead arrives there as cells progress through the cell cycle, which presumably enables it to be present at newly formed

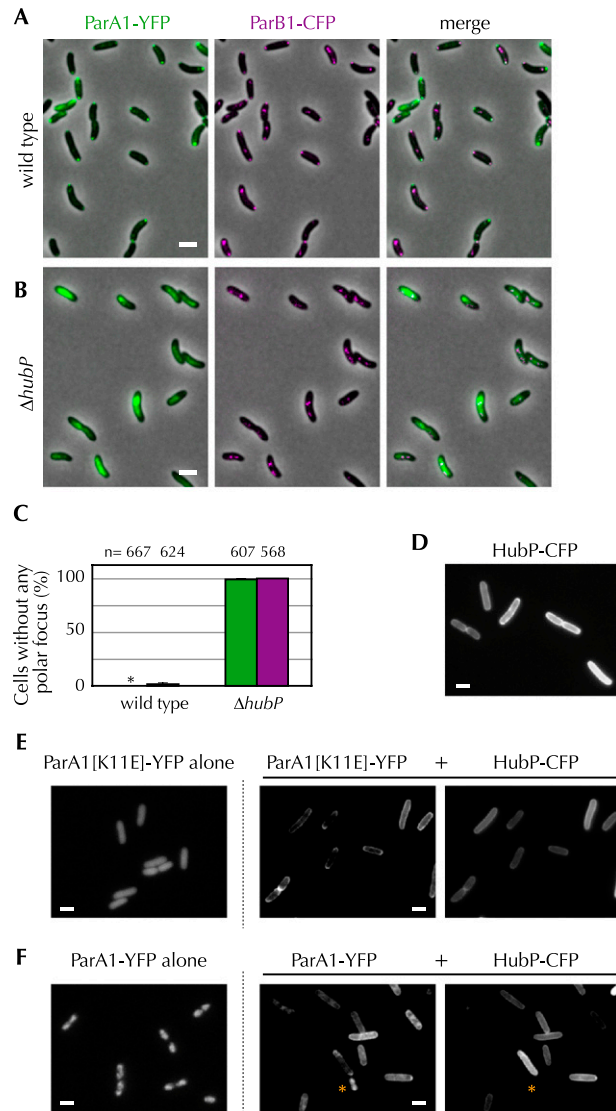


poles (Fig. 2C). HubP localization was similar using plasmid- and chromosome-encoded protein, although the chromosome-encoded protein was less abundant (Supplemental Movies S1, S2).

Fluorescence microscopy was also used to explore the dynamics and determinants of HubP's subcellular localization. Notably, photobleaching of polar HubP-GFP was followed by recovery of the polar signal, revealing that HubP can be directly targeted to the pole; it does not need to arrive there via the cell division site. Recovery of fluorescence intensity at the bleached pole was accompanied by a decline of fluorescence intensity at the opposite (unbleached) pole, suggesting that there is an exchange of HubP between the two poles (Fig. 2F). Additionally, we found that polar targeting of HubP is independent of its putative cytoplasmic and membrane-spanning domains. A fusion protein consisting of the first 324 amino acids of HubP (the putative periplasmic and transmembrane sequences) fused to CFP had a cellular distribution equal to that of the full-length fusion protein (Fig. 2G). A smaller (1–284) fragment fused to CFP was not detected, consistent with the predicted localization of the HubP N-terminal domain in the periplasm (Fig. 2A), where CFP does not fluoresce. However, when smaller N-terminal fragments were fused to mCherry (which is fluorescent even in the periplasm), polar targeting was also observed (Fig. 2G, 1–284). The first 161 amino acids of HubP were sufficient to enable polar targeting of the fusion protein, although the first 150 amino acids were not (Fig. 2G, 1–161 and 1–150). We also found that polar targeting of HubP required the putative LysM domain (amino acids 90–134) (Fig. 2G,  $\Delta$ LysM). In the absence of targeting sequences, HubP appeared to be diffusely distributed in the periplasm (Fig. 2G, 1–150 and  $\Delta$ LysM). The requirement for the LysM domain raises the possibility that an interaction between HubP and PG contributes to its targeting and/or retention at the *V. cholerae* pole. However, to date, we have been unable to detect a direct interaction between HubP and PG using in vitro binding assays.

#### HubP modulates the localization of *oriCI*

We explored whether the failure of ParA1 to localize to the cell poles in cells lacking HubP altered *chrI* segregation dynamics. Although ParA1 is not essential for *chrI* partitioning, in its absence, *oriCI* and the origin-associated centromere-binding protein ParB1 do not localize to the old pole in young cells, nor are they drawn to the new pole following chromosome replication (Fogel and Waldor 2006). Instead, in a *parA1* deletion mutant, they are generally found at the mid-cell of young cells and near the 1/4 and 3/4 positions following replication (Fogel and Waldor 2006). We found that deletion of *hubP* has a similar effect on the localization of ParB1: No polar ParB1-CFP foci were detected in the *hubP* mutant, and cells typically contained two well-separated cytoplasmic foci (Fig. 3A–C), as observed in *parA1* mutants. Thus, marking of the *V. cholerae* cell pole by HubP is critical for the establishment of the normal cellular distribution of the organism's major chromosome.



**Figure 3.** HubP governs the subcellular distribution of ParA1. (A–C) Subcellular localization of ParA1-YFP and ParB1-CFP in wild type (A) and  $\Delta$ hubP (B) *V. cholerae* cells. (C) The percentage of wild-type and  $\Delta$ hubP cells lacking polar foci of ParA1 (green) and ParB1 (magenta) is shown, based on analysis of the number of cells (*n*) indicated above. Asterisk indicates 0%. (D) Fluorescence of HubP-CFP expressed in *E. coli*. (E,F) ParA1[K11E]-YFP (E) or ParA1-YFP (F) was expressed in *E. coli* in the absence (left) or presence (middle) of HubP-CFP (right). ParA1-YFP localized to the nucleoid in the cell lacking HubP-CFP expression (marked with an asterisk). Representative fields are shown. Bar, 2  $\mu$ m.

#### HubP binds directly to ParA1 to control its subcellular distribution

Several lines of evidence suggest that HubP controls ParA1's distribution via a direct interaction between the two proteins. First, when ParA1[K11E]-YFP and HubP-CFP were coexpressed in *V. cholerae*, the foci they formed colocalized (Fig. 2D; see below). Second, these proteins were found to interact in a bacterial two-hybrid assay, which also showed that ParA1 can self-associate (Table 1;

**Table 1.** Summary of HubP's interactions

Protein	Experimental approach	HubP region		
		Full-length	[1–1440]	[1–900]
ParA1	BACTH <sup>a,b</sup>	++	+	–
	FP <sup>c</sup> in <i>E. coli</i> <sup>d</sup>	+	+	–
	FP in <i>V. cholerae</i> <sup>c</sup>	+	+	–
ParA1 [K11E]	BACTH <sup>b</sup>	–	— <sup>f</sup>	— <sup>f</sup>
	FP in <i>E. coli</i> <sup>d</sup>	+	+	–
	FP in <i>V. cholerae</i> <sup>c</sup>	+	+	–
ParC	BACTH <sup>b</sup>	–	— <sup>f</sup>	— <sup>f</sup>
	FP in <i>E. coli</i> <sup>d</sup>	— <sup>g</sup>	— <sup>g</sup>	–
	FP in <i>V. cholerae</i> <sup>c</sup>	+	+	+
FlhG	BACTH <sup>b</sup>	+	+/–	+/–
	FP in <i>E. coli</i> <sup>d</sup>	+	— <sup>g</sup>	–
	FP in <i>V. cholerae</i> <sup>c</sup>	+	–	–
FlhF	BACTH <sup>b</sup>	+++	–	–
	FP in <i>E. coli</i> <sup>d</sup>	+	–	–
	FP in <i>V. cholerae</i> <sup>c,h</sup>	+	— <sup>f</sup>	— <sup>f</sup>

<sup>a</sup>(BACTH) Bacterial two-hybrid.<sup>b</sup>See also Supplemental Figure S3.<sup>c</sup>(FP) Colocalization of fluorescent proteins.<sup>d</sup>See also Supplemental Figure S5.<sup>e</sup>See also Figure 6.<sup>f</sup>Not tested.<sup>g</sup>Ambiguous result.<sup>h</sup>Note that FlhF forms polar foci even in the  $\Delta hubP$  background.

Supplemental Fig. S3), similar to observations for other ParA-like proteins (Leonard et al. 2005; Ebersbach et al. 2006). Finally, HubP recruited ParA1 to the cell periphery when the two proteins were coexpressed in *Escherichia coli* (Fig. 3D–F). When expressed alone in *E. coli*, ParA1[K11E]-YFP and ParA1-YFP were widely distributed within the cytoplasm or associated with the nucleoid, respectively (Fig. 3E,F). However, when expressed along with HubP-CFP, which appears to be membrane-associated but not polar in *E. coli* (Fig. 3D), the distribution of both wild-type ParA1 and ParA1[K11E] was shifted toward the cell membrane (Fig. 3E,F).

A variety of mutations within *parA1* was generated in order to explore which features of the protein are needed for interaction with HubP. Based on previous analyses of ParA family members and identification of key conserved amino acids, mutations were generated to disrupt dimerization (ParA1[G12V]), DNA binding (ParA1[R189E]), ATP binding (ParA1[K16A]), and ATP hydrolysis (ParA1[D40A]), and the subcellular distribution of the mutant proteins was analyzed in *E. coli* and *V. cholerae*. Mutants predicted to be unable to dimerize or bind DNA still appeared capable of interacting with HubP in both systems; they formed HubP-dependent polar foci in *V. cholerae* and showed HubP-dependent membrane association in *E. coli* (Supplemental Fig. S4). Mutants lacking the ability to bind or hydrolyze ATP also were recruited by HubP to the cell periphery in *E. coli* and formed HubP-dependent polar foci in *V. cholerae* (Supplemental Fig. S4). However, nonpolar ParA1[K16A] and ParA1[D40A] foci were also detected in both wild-type and *hubP* cells regardless of the presence or absence of a chromosomal *parA1*<sup>+</sup> copy, suggesting that

ParA1 binding to and/or hydrolysis of ATP is not essential for, but may influence, ParA1–HubP interactions.

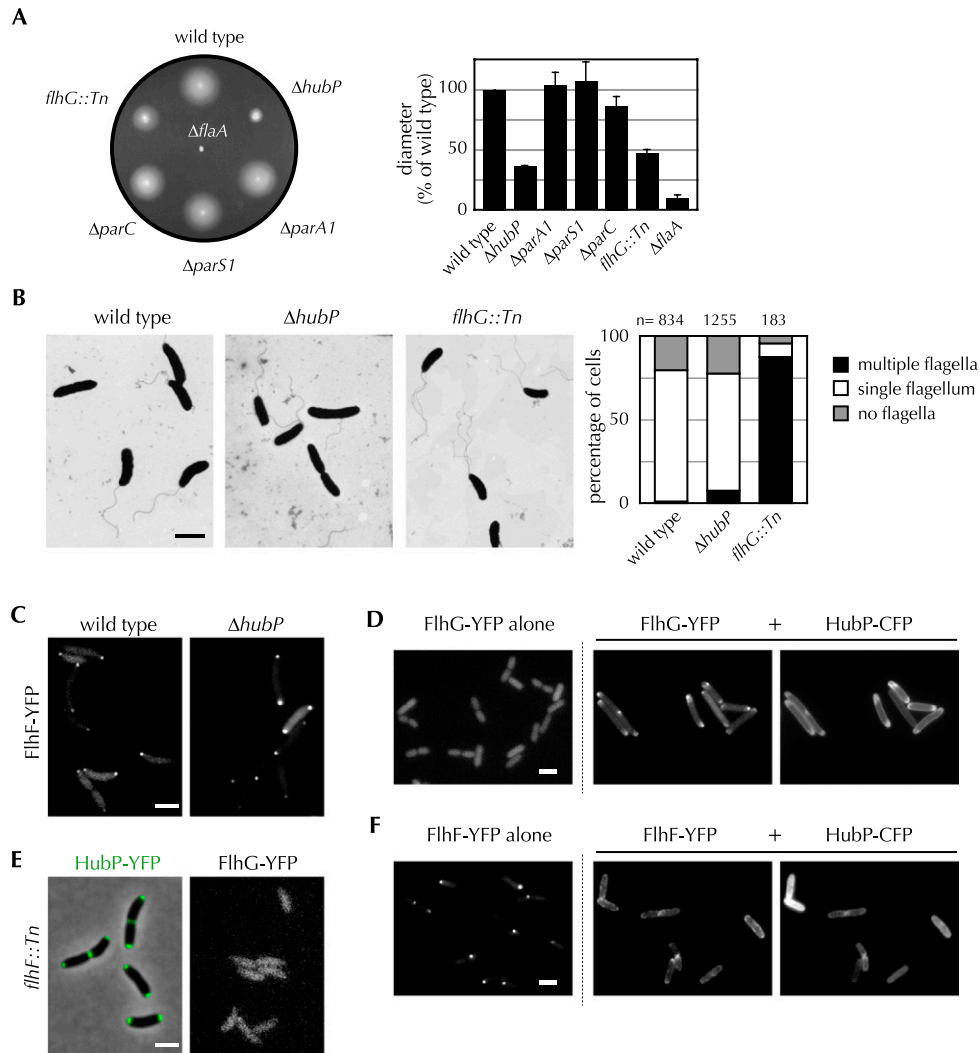
### *HubP negatively regulates polar accumulation of flagella*

Our observation of FlhG mislocalization in the *hubP* mutant, coupled with the previously noted reduced motility of this mutant on soft agar plates, led us to further explore the connection between HubP and *V. cholerae* flagellation. The *hubP* mutant exhibited a >50% reduction in swarming on soft agar, comparable with the defect displayed by an *flhG* mutant and significantly larger than that of a *parC* mutant (Fig. 4A). However, deletion of *hubP* had a fairly modest effect on flagellation. Electron microscopy analyses revealed that a slightly increased fraction (6%) of *V. cholerae* lacking HubP produces more than a single polar flagellum, a feature observed in only 1% of wild-type cells (Fig. 4B). This frequency contrasts markedly with that of an *flhG* mutant, where ~80% of cells had multiple flagella (Fig. 4B). In *Vibrio alginolyticus*, FlhG is thought to repress flagellar formation via two mechanisms: (1) by binding to FlhF (which is thought to guide polar placement of flagella) and preventing its polar recruitment, and (2) by repressing expression of flagellar genes (Kusumoto et al. 2008). In *V. cholerae*, deletion of *hubP* does not have a significant effect on the distribution of FlhF, despite its effect on FlhG localization, suggesting that polar FlhG is not a critical determinant of FlhF accumulation at this site (Fig. 4C). Furthermore, the relatively minor effect of FlhG mislocalization on flagella production in the *hubP* mutant suggests that mislocalization does not substantially impair its efficacy in negatively regulating flagellar production. Thus, the biological rationale for polar targeting of FlhG by HubP remains to be determined.

### *HubP interacts directly with FlhG and FlhF*

Like ParA1, FlhG appears to be retained at the *V. cholerae* pole through direct interaction with HubP. Bacterial two-hybrid analyses suggest that the two proteins interact (Table 1; Supplemental Fig. S3), and coexpression of HubP along with FlhG in *E. coli* shifts FlhG from a diffuse cytoplasmic distribution to a pole-associated distribution that corresponds to the location of HubP (Fig. 4D). It is not clear why coexpression of these two proteins also alters the subcellular distribution of HubP (cf. Figs. 3D–F and 4D).

Unexpectedly, two-hybrid analyses suggest that FlhF also interacts with HubP (Table 1), although neither protein is required for the polar targeting of the other (Fig. 4C,E). Analyses of FlhF and HubP localization in *E. coli* also suggest that they interact; expression of HubP shifts FlhF from a predominantly polar distribution to a pattern that matches HubP (Fig. 4F). Notably, FlhF, like HubP, is required for recruitment of FlhG to the *V. cholerae* pole (Fig. 4E). Thus, although the precise reason for an interaction between FlhF and HubP is unknown, it is possible that they act together to modulate the subcellular distribution of FlhG.



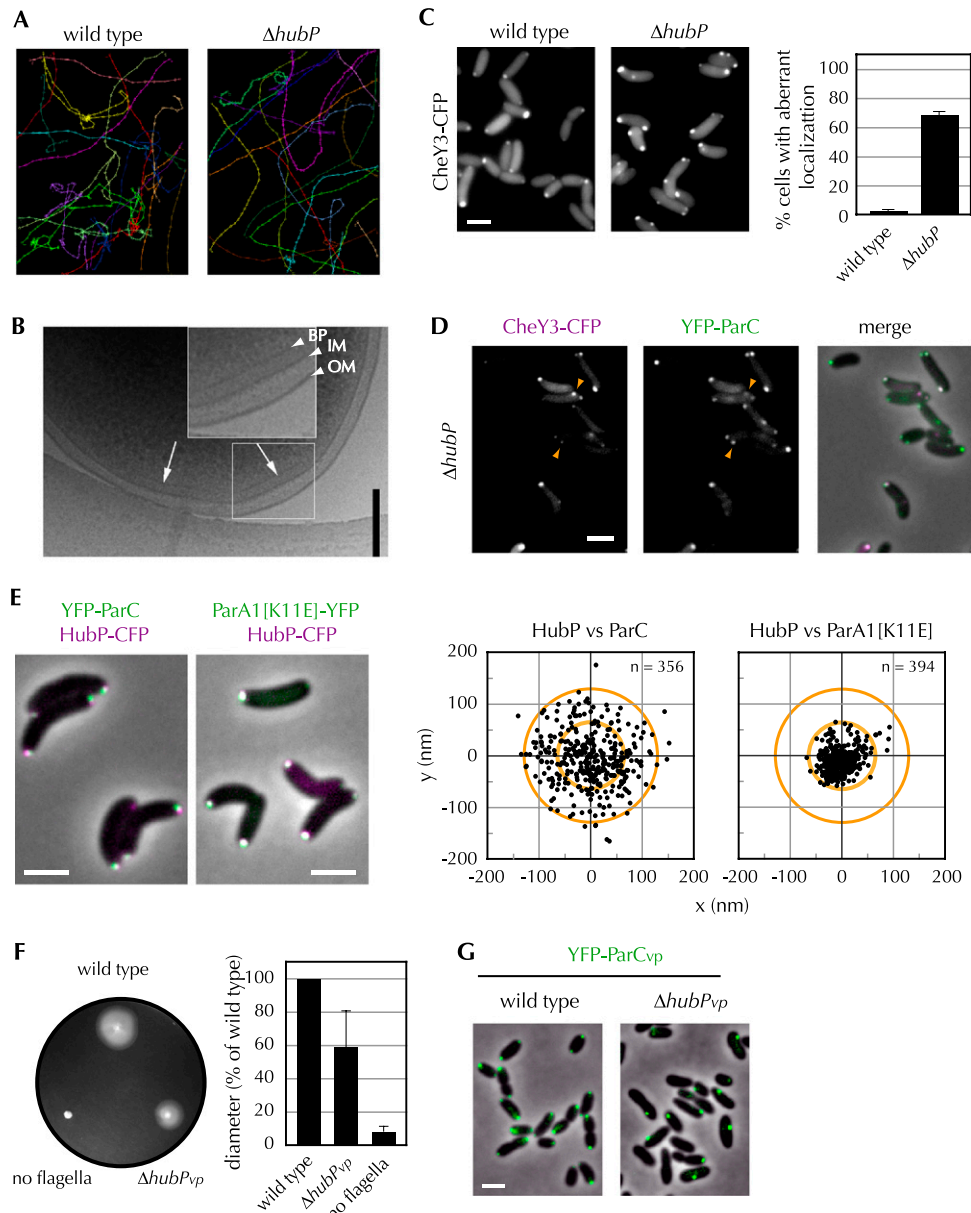
**Figure 4.** HubP is important for *V. cholerae* chemotaxis and regulation of flagellar production. (A) Swarming of various *V. cholerae* mutants on a representative soft agar plate is pictured (left), and average swarm diameter relative to wild type, with standard deviation based on three or more experiments, is graphed (right). (B, left) Images of wild-type, *hubP*, and *flhG* *V. cholerae*, generated using electron microscopy, are shown. The number of flagella observed per cell for each mutant is graphed at the right. (C,E) Subcellular localization of FlhF-YFP (C), HubP-YFP (E), and FlhG-YFP (E) in the indicated *V. cholerae* strains. (D,F) FlhG-YFP (D) or FlhF-YFP (F) was expressed in *E. coli* in the absence (left) or presence (middle) of HubP-CFP (right). Representative fields are shown. Bar, 2  $\mu$ m.

#### Deletion of *hubP* impairs polar localization of chemotactic signaling proteins and polar assembly of chemoreceptor arrays

In contrast to its relatively minor effect on flagellation, deletion of *hubP* has a far more dramatic effect on *V. cholerae* chemotaxis. Video tracking of swimming bacteria revealed that the *hubP* strain displays a significant bias toward straight swimming: Direction reversals were detected  $0.16 \pm 0.07 \text{ sec}^{-1}$  for the mutant versus  $0.57 \pm 0.06 \text{ sec}^{-1}$  for the wild-type strain (Fig. 5A). The effect of *hubP* deletion on swimming is more marked than that of a *parC* deletion ( $0.28 \text{ sec}^{-1}$ ), consistent with its more pronounced effect on swarming on soft agar plates (Fig. 4A). Chemotaxis appears to be impaired due to mislocalization of components of chemotactic apparatus in the

*hubP* mutant. As previously shown (Briegleb et al. 2009), electron cryotomography revealed that 39 of 100 wild-type cell poles examined had detectable chemoreceptor arrays (Fig. 5B). In marked contrast, only seven of 61 (11%) *hubP* cell poles had detectable arrays. Consistent with the reduced number of polar arrays, chemotaxis signaling protein CheY3 formed aberrant nonpolar foci in ~60% of *hubP* cells (Fig. 5C). Furthermore, most of the nonpolar CheY3 foci colocalized with ParC (Fig. 5D), which we showed is also mislocalized in the *hubP* mutant (Fig. 1C). Together, these observations suggest that HubP (via ParC) promotes the polar localization of the entire chemoreceptor/signaling complex. Thus, at least in *V. cholerae*, the distribution of receptors and downstream signaling proteins is not simply the result of stochastic processes.

## A multidomain hub organizes the cell pole



**Figure 5.** Deletion of *hubP* indirectly alters the distribution and efficacy of vibrio chemotaxis proteins. (A) Positional tracking of swimming wild-type and  $\Delta hubP$  *V. cholerae*. (B) Electron cryotomographs of wild-type *V. cholerae*. The region in the square is magnified in the *inset*. The arrow indicates chemoreceptor array. Arrowheads indicate outer membrane (OM), inner membrane (IM), and base plate (BP) of chemoreceptor array. (C) Foci of CheY3-CFP in wild-type and  $\Delta hubP$  *V. cholerae* (left) and percentage of cells with mislocalized CheY3-CFP in indicated background (right). (D) CheY3-CFP and YFP-ParC foci in  $\Delta hubP$  *V. cholerae*. Arrowheads indicate colocalization of nonpolar CheY3-CFP and YFP-ParC foci. (E) HubP-CFP (magenta) colocalizes with ParA1[K11E]-YFP (right; green) but not with YFP-ParC (left; green). Localization of YFP-ParC and ParA1[K11E]-YFP foci relative to neighboring HubP-CFP focus are plotted. Inner and outer circles correspond to diameters of 2 and 4 pixels, respectively. (F) Swarming of wild-type,  $\Delta hubP$ , and flagella mutant (lacks both polar and lateral flagella) *V. parahaemolyticus* on a representative soft agar plate is pictured (left), and average swarm diameter relative to wild type, with standard deviation based on three or more experiments, is graphed (right). (G) Subcellular localization of YFP-ParC<sub>vp</sub> in the indicated *V. parahaemolyticus* strain. Representative fields are shown. Bar, 2  $\mu$ m.

In contrast to ParA1 and FlhG, ParC does not appear to be anchored at the pole by directly interacting with HubP. The two proteins did not interact in a two-hybrid assay (Table 1); furthermore, there was at most a subtle change in the distribution of YFP-ParC in *E. coli* induced by coexpression of HubP-CFP, and the two proteins do not

appear to be colocalized (Supplemental Fig. S5). Finally, although HubP-CFP and YFP-ParC are both polar proteins in *V. cholerae*, they do not precisely colocalize. Mapping of ParC's position relative to HubP (and ParA1[K11E]'s, for comparison) revealed that ParC was, on average, approximately two times farther from HubP than was



ParA1[K11E]:  $68.4 \pm 40.4$  nm versus  $30.9 \pm 16.4$  nm (Student's *t*-test:  $1.2 \times 10^{-54}$ ) (Fig. 5E). Collectively, these observations suggest the possibility that HubP modulates the distribution of ParC indirectly, perhaps via the mediation of an as-yet-undefined vibrio-specific factor. However, ParA and the other ParA-related ATPases do not serve this function, as previous analyses have shown that ParC localization is independent of ParA1 and FlhG as well as of FlhF (Ringgaard et al. 2011).

#### *HubP promotes the polar localization of ParC and ParA in V. parahaemolyticus*

Since the aberrant chemotaxis of the *V. cholerae* *hubP* mutant (Figs. 4A, 5A) was a particularly striking phenotype, we wondered whether *hubP* also modulates chemotaxis in other vibrios (e.g., *V. parahaemolyticus*). Like all vibrios and photobacteria, *V. parahaemolyticus* encodes homologs of *V. cholerae* HubP and ParC (69% and 86% similarity, respectively). We found that *V. parahaemolyticus* swarming motility, which is dependent on the organism's polar flagella, was reduced by ~40% in a *V. parahaemolyticus* *hubP* mutant (Fig. 5F). Furthermore, as in *V. cholerae*, YFP-ParC routinely formed polar foci in wild-type *V. parahaemolyticus*. In the *hubP* mutant, however, YFP-ParC was often observed within nonpolar foci as well as polar foci (Fig. 5G). Thus, it seems likely that HubP's role as an organizer of chemotactic machinery is conserved, at least within the *Vibrionaceae*. Interestingly, there is a striking correlation in the coincidence of *hubP* and *parC* homologs among  $\gamma$ -proteobacteria (Supplemental Table S1), suggesting that HubP's influence over chemotaxis may extend through a broader swath of the bacteria kingdom. There is much less overlap between the distribution of HubP and ParA homologs (which are far more abundant). Nonetheless, we observed that HubP also appears to mediate the polar localization of ParA1 in *V. parahaemolyticus* (data not shown), indicating that HubP's role in chromosome partitioning is likewise not limited to *V. cholerae*.

#### *Different HubP domains are required to mediate polar localization of ParA1, FlhG, and ParC*

Our observation that HubP interacts directly with ParA1 and FlhG, but not with ParC, suggested that distinct regions of HubP might be important for determining the subcellular distribution of client proteins. We explored this possibility using the bacterial two-hybrid system and the fluorescence-based assays described above, except that *hubP* was altered by the presence of a series of deletion mutations (summarized in Table 1; data presented in Fig. 6; Supplemental Figs. S3, S5). Results from all of these systems suggest that the interaction between ParA1 and HubP requires amino acids within the repeat region of HubP. Truncation of the protein to remove the final 181 amino acids did not impair HubP's ability to interact with or guide the localization of ParA1 (*hubP*[1–1440]) (Fig. 6A,B); however, deletion of the repeats (*hubP*[0-repeat]) or of the repeats along with the C terminus (*hubP*[1–900]) eliminated HubP's activity with respect to ParA1 (and ParA1[K11E]). In

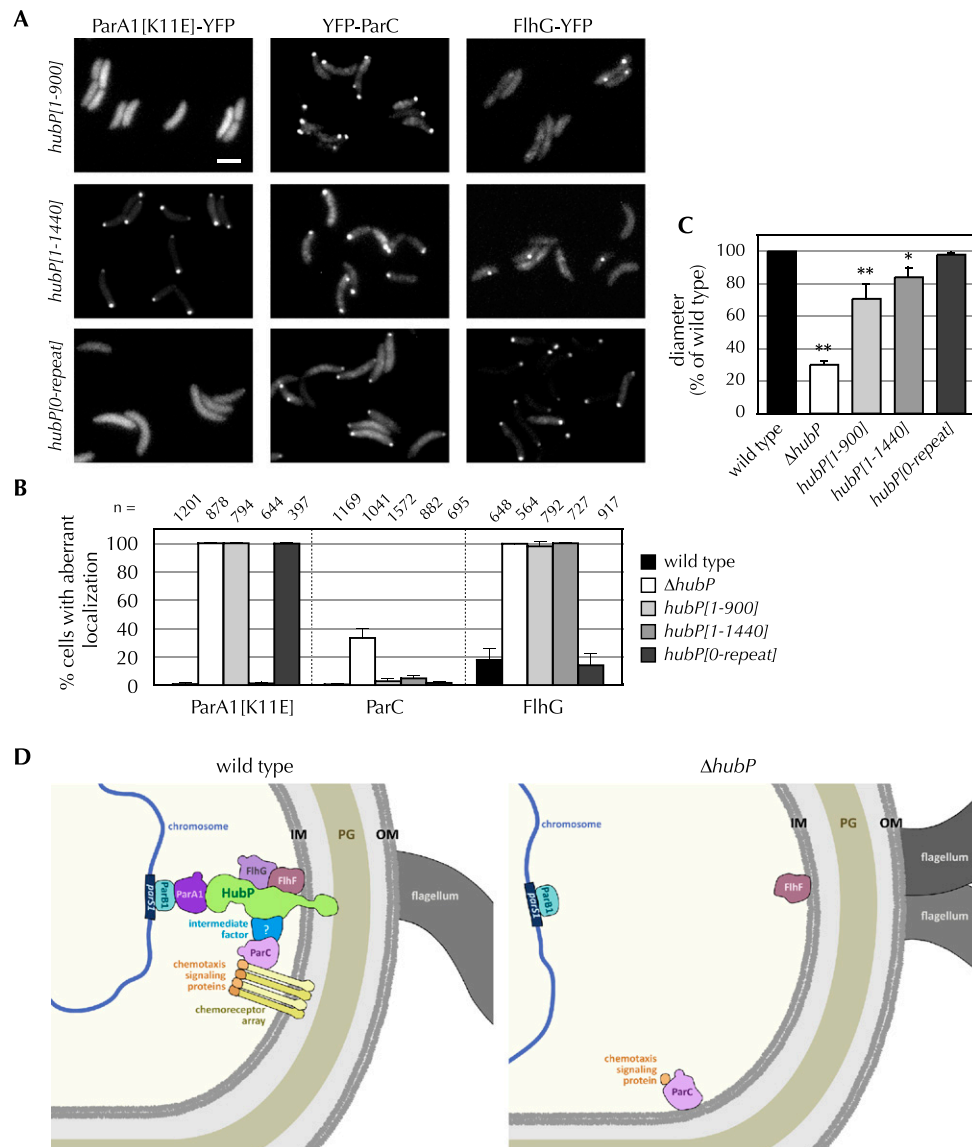
contrast, the extreme C terminus of HubP (amino acids 1441–1621) appeared to be essential for interaction between HubP and FlhG, at least based on FlhG-YFP localization in *V. cholerae* expressing HubP from the chromosome (Fig. 6A,B). Plasmid-based assays in *E. coli* suggest that additional regions of HubP may have a weak capacity to interact with FlhG (Supplemental Fig. S3); nonetheless, it is clear that amino acids 1441–1621 are important using these other assays as well. The C-terminal 181 amino acids were also critical for the interaction between HubP and FlhF (Supplemental Fig. S5F). Finally, HubP's modulation of ParC localization appears to depend on yet another domain within HubP. The first 900 amino acids of HubP were sufficient to localize ParC normally in *V. cholerae* (Fig. 6B), and chemotaxis (soft agar motility) by *V. cholerae* expressing HubP[1–900] was significantly greater than by a strain lacking the entire protein (Fig. 6C). Chemotaxis by the *hubP*[1–900] mutant was not equivalent to that by the wild-type strain; however, this may reflect the inadequacy of the truncated HubP for modulation of flagellar proteins. Consistent with this hypothesis, HubP[0-repeat], which interacts with both ParC and FlhG, enabled chemotaxis equivalent to that of the wild-type strain. Collectively, these results suggest that the cytoplasmic portion of HubP is likely to have multiple distinct interaction domains that mediate its spatial regulation of ParA family proteins.

## Discussion

In rod-shaped bacteria, the cell poles are critical subcellular domains, but knowledge of the mechanisms that underlie pole formation, recognition, organization, and perpetuation is restricted to very few model organisms. In *V. cholerae*, polarly localized ParA-related proteins are essential for mediating the polar localization of DNA/protein (ParA1 and *parS1*/ParB1) and protein/protein complexes (ParC and CheY3) that are required for proper segregation of *oriCI* and the chemotactic machinery, respectively. FlhG, a third polar ParA-like protein, modulates formation of *V. cholerae*'s polar flagellum. Here, we found that the polar localization of ParA1, ParC, and FlhG all depend on HubP, a large transmembrane protein that is conserved in all *Vibrionaceae*/*Photobacteriaceae* as well as in several other  $\gamma$ -proteobacteria. In the absence of this polar anchor, origin segregation and chemotaxis are impaired, and cells produce extra flagella. Polar targeting of ParA1, ParC, and FlhG requires different regions within HubP, and the ParA family proteins clearly use distinct mechanisms (including both direct and indirect interactions with HubP) for their polar tethering. Thus, the organization of the vibrio cell pole is dependent on a complex and multifaceted protein hub that anchors chromosome segregation and chemotactic machinery to this site (Fig. 6D).

Strikingly, although HubP is required for the polar placement of its ParA-related clients, there is not a perfect correspondence between their subcellular localizations. HubP is always detectable at both the old and new pole in *V. cholerae* and often at the future division plane at mid-cell as well. In contrast, ParC does not form detectable

## A multidomain hub organizes the cell pole



**Figure 6.** HubP appears to have multiple domains for interaction with partners. (A) Fluorescent fusion proteins (as indicated) were visualized in *V. cholerae* in which full-length *hubP* was replaced by a truncated gene. Representative fields are shown. Bar, 2  $\mu$ m. (B) For each protein indicated, the frequency with which the aberrantly localized fusion protein was observed is plotted as a function of *hubP* genotype. Protein localization was scored in the indicated number (*n*) of cells. For ParA1 and FlhG, aberrant localization refers to cells lacking a polar focus; for ParC, it reflects the number of cells with a nonpolar focus. Mean and standard deviations and total number of cells counted (*n*) are shown. Wild-type and  $\Delta hubP$  data are reiterated from Figure 1E for comparison. (C) Swarming of *V. cholerae* *hubP* mutants on soft agar plates was quantitated, relative to wild type, based on swarm diameter. Truncated HubP (as indicated) was expressed from an allele inserted within the chromosome in place of the wild-type gene. Plotted values reflect the means and standard deviations from three or more experiments. (\*)  $P < 0.05$ ; (\*\*)  $P < 0.01$  by Student's *t*-test. (D) Schematic of HubP's role in organizing the *V. cholerae* old cell pole. In wild-type cells, different regions of HubP anchor three ParA-related ATPases (ParA1, ParC, and FlhG) to the old cell pole. These ATPases mediate the polar localization of the centromeric ParB1/*parS*1 complex and chemotaxis receptor arrays and signaling proteins, respectively. HubP anchoring of ParC to the pole appears to be mediated through an intermediate factor. In the absence of HubP ( $\Delta hubP$ ), ParA1 and FlhG are diffuse, the ParB1/*parS*1 complex is no longer polar, ParC and chemotaxis signaling proteins form nonpolar foci (although polar foci are observed as well), and there is a small increase in cells that contain two flagella; FlhF, which also interacts with HubP, remains at the pole.

foci at the new pole until late in the cell cycle (Ringgaard et al. 2011), and FlhG is only bipolar in a small subset of (typically long) cells (Fig. 1). Distinct ParA1 foci are also difficult to discern at the new pole until late in the cell

cycle, which corresponds to the migration of replicated *oriC*1 to this site. However, it is presumed that some ParA1 must be anchored at the new pole at the start of chromosome segregation in order for cycles of ParA1

polymerization/depolymerization to yield a net force pulling ParB1/*oriCI* across the cell (Fogel and Waldor 2006). Nonetheless, there appear to be restrictions on interactions between HubP and its clients at the new pole (and at the mid-cell prior to division). What limits HubP interactions with its clients at the new pole? It is possible that additional factors must be present at this site along with HubP in order to anchor HubP's partners (as appears to be the case for ParC). Alternatively, interactions at the new pole may be limited by the presence of an additional factor that is released or inactivated later in the cell cycle. Client-specific processes, such as interactions with additional protein partners, ATP, or the nucleoid, may also regulate protein localization and may account for the apparently distinct arrival times of ParA1, FlhG, and ParC at the new pole. It remains to be seen whether there is any temporal coupling among these processes (e.g., between *oriCI* segregation and movement of ParC to the new pole) or whether they are fully independent.

Similarly, it is noteworthy that some HubP-dependent proteins are not exclusively targeted to either pole for at least a portion of the cell cycle. Much of ParA1 is present within the cytoplasm, although its distribution there is dynamic, and much of FlhG is diffusely localized. Polar versus cytoplasmic localization is likely modulated at least in part by ATP binding and hydrolysis. Binding of ParA-like proteins to ATP typically enables them to dimerize, and dimerization is often required for their interactions with both other proteins (e.g., ParB and homologs) and DNA (Lutkenhaus 2012). Interestingly, our analyses indicate that ParA1 dimerization is not required for its interaction with HubP. In fact, ParA1[G12V], which is thought to not dimerize, is even routinely found at new poles (Supplemental Fig. S4), suggesting that dimerization may inhibit interaction with HubP. Our analyses also suggest that ATP binding and hydrolysis are not essential for the interaction between ParA1 and HubP, since ParA1[K16A] and ParA1[D40A], which fail to bind and hydrolyze ATP, respectively, still form some polar foci, although they do have a reduced presence at the poles relative to wild-type ParA1. The ability of client proteins to bind ATP could also indirectly modulate interactions with HubP; e.g., by altering their affinity for and positioning by other interaction partners. Ultimately, the distribution of HubP's clients between the cytoplasm and the pole likely reflects the relative affinities of the client proteins for their cytoplasmic partners versus HubP.

The means by which nascent HubP is initially recruited to the mid-cell or poles warrants further investigation. Polar targeting of HubP could be due to recognition of membrane curvature, as has been proposed for DivIVA of *Bacillus subtilis* (Lenarcic et al. 2009; Ramamurthi and Losick 2009). However, our preliminary analyses of HubP's distribution in the spherical or oval cells formed due to mutation of *rodZ* (Shiomi et al. 2008; Alyahya et al. 2009; Bendezú et al. 2009) and in MreB inhibitor MP265-treated (Takacs et al. 2010) cells suggest that HubP may not simply recognize the membrane curvature associated with polar regions, as HubP retains a focal distribution even in these cases. One possibility is that HubP recognizes features of

PG that are stable and specific to the poles. Polar PG is thought to be relatively inert, and pole-specific PG structures formed during cell division might serve as a "birth scar" that might be used in proper cellular targeting of HubP. There is a precedent for PG fastening other proteins to poles; e.g., *C. crescentus* SpmX (Radhakrishnan et al. 2008). Alternatively, HubP association with polar PG (presumably through its LysM domain) may only stabilize its association with the poles after it is recruited there by an unknown factor. Determination of the relative timing of PG synthesis at the mid-cell versus HubP recruitment there could aid in distinguishing between these possibilities.

Although HubP routinely localizes at the poles and may interact with polar PG, HubP is not stably retained at these sites; instead, it can migrate from one pole to the other, apparently bidirectionally. The significance of and mechanism underlying this movement, which has not been reported for other polar markers, remain to be determined. Additionally, it has not been investigated whether partner proteins remain associated and move with HubP or whether they are instead released and recaptured. However, the fact that HubP is detected at the mid-cell or new pole of young cells apparently without associated partners may indicate that partners are released prior to or during movement. It is also possible that HubP acquires interacting partners during its transits across the cell. If so, bipolar movement of HubP might serve as a tool for recruiting its interaction partners to polar sites.

The most profound consequences of *hubP* deletion that we observed relate to its regulation of chemotaxis. In the absence of HubP, polar chemotactic receptor arrays are present at a markedly reduced frequency, and chemotactic signaling complexes (marked by ParC and CheY3) are often mislocalized, although they are assumed to be structurally intact. The altered distribution of these proteins presumably is the major factor underlying the mutant's impaired chemotaxis, detectable as reduced swarming on soft agar plates and a reduced reversal frequency in liquid culture. Notably, a HubP deficiency has a more profound phenotype than does a ParC deficiency, probably both because ParC does not modulate flagellar assembly and because the frequency at which receptors and associated signaling proteins accumulate at the pole remains relatively high in the absence of ParC, so signaling is not perturbed. In contrast, in the absence of HubP, ParC is frequently mislocalized, causing mislocalization of downstream signaling proteins. However, downstream signaling proteins are also frequently mislocalized in a strain lacking *hubP* and *parC* (data not shown). Thus, although mislocalization of ParC may be more detrimental to *V. cholerae* than the total absence of this protein, it is possible that ParC is not the only factor through which HubP controls the localization of chemotaxis proteins. Contrasting paradigms characterize HubP's impact on FlhG and ParA1. Mislocalization of FlhG has a relatively minor effect on flagellar assembly, far less than that of FlhG's absence, perhaps because some of FlhG's regulatory roles are not dependent on its subcellular distribution. Finally, the mislocalization and absence of ParA1 have similar consequences; both result in aberrant subcellular placement of *oriCI*.

Although the precise means by which HubP recruits and/or anchors ParC to the pole and thereby situates the remainder of the chemotactic machinery remains to be determined, bioinformatic analyses suggest that it may be a relatively conserved process. There is a strong correlation between genomes containing *hubP* and genomes containing *parC*, particularly among the  $\gamma$ -proteobacteria (Supplemental Table S1). Notably, since our data suggest that ParC and HubP do not interact directly, it is likely that an additional component that serves as a bridge between these two proteins will also be found to have coevolved.

HubP bears some functional similarity to TipN and PopZ, polar determinants and markers in *C. crescentus*, the organism in which the molecular bases for pole establishment and organization are best understood. Through their presence at the poles, all three of these proteins enable polar placement/anchoring of several additional proteins and protein complexes, such as the chromosome segregation machinery, whose proper activity depends on their polar localization. TipN, like HubP, is an early marker of new/nascent poles; both proteins are present at the mid-cell prior to cell division, thereby providing cells with cues regarding polar orientation at the start of their development. However, there are also notable differences in how these three pole-specifying proteins localize and function. In particular, HubP differs from TipN and PopZ in that it is always present at both poles, rather than marking the new pole (TipN) or primarily the old pole (PopZ). It seems possible that additional factors/processes in *V. cholerae* are therefore needed for the cell to distinguish between its polar regions. Additionally, there is no evidence that HubP is a primary determinant, rather than a marker, of pole formation, unlike TipN, whose overexpression can induce ectopic pole formation (Lam et al. 2006). Furthermore, HubP's clients identified to date are limited to ParA-like ATPases, whereas TipN and PopZ modulate the localization of structurally diverse proteins, although these include ParA family members (Schofield et al. 2010). The *C. crescentus* and *V. cholerae* pole-organizing proteins lack amino acid sequence similarity and clearly evolved through independent processes in  $\alpha$ -proteobacteria and  $\gamma$ -proteobacteria. Despite their differences, it is striking that their core characteristics—polar proteins that anchor multiple clients to generate and perpetuate cell polarity—have so much in common. It seems likely that additional diverse hub proteins will control the activity of the growing family of ParA-related proteins that govern the polar localization of numerous subcellular structures.

## Materials and methods

### Plasmids and strains

Plasmids used in this study and their construction are described in the Supplemental Material (Supplemental Table S2; Supplemental Material). Oligonucleotides used for plasmid construction are listed in Supplemental Table S3. Nucleotide sequences of plasmids were confirmed whenever the construction involved

PCR. Strains used in this study are listed in Supplemental Table S4. Briefly, we used El Tor O1 strains N16961 (Heidelberg et al. 2000) and C6706 (parental strain of the mapped Tn insertion library) (Cameron et al. 2008) as wild-type *V. cholerae*. Chromosomal deletions and insertions were introduced by allelic exchange (Donnenberg and Kaper 1991). For growth conditions and supplements including antibiotics, see the Supplemental Material.

### Screening

The mapped Tn insertion library of *V. cholerae* contains 147 mutants showing a motility deficiency (<50% compared with wild type) (Cameron et al. 2008). Of these 147 mutants, the 56 that have Tn insertions in genes obviously linked to motility (e.g., flagellar biosynthesis genes) were discarded from the pool of candidates. A reporter plasmid encoding fluorescent fusion proteins, pBAD33 parA1[K11E]-yfp cheY3-cfp, was introduced into the remaining 91 mutants, and the subcellular localization of the fusion proteins in log-phase cells was manually examined using fluorescence microscopy.

### Microscopy

For microscopy, see the Supplemental Material.

### Electron microscopy

Cells were grown to O.D. ~0.5 in LB, immobilized on grids, fixed with glutaraldehyde, and then repeatedly washed in water. Imaging was performed on a Tecnai G<sup>2</sup> Spirit BioTWIN.

### Cryomicroscopy

For cryomicroscopy, see the Supplemental Material.

## Acknowledgments

We are grateful to Janet Iwasa for making Figure 6D, Paula Montero Llopis for an expert tutorial in Microbe Tracker, and John Mekalanos for providing the mapped transposon library. We also thank Richard Losick, Hubert Lam, and Waldor laboratory members for helpful discussions and comments on the manuscript. S.R. was funded with a post-doctoral fellowship from the Villum Kann Rasmussen Foundation. This work was supported by the NIAID R37 AI-042347 (to M.K.W.), NIGMS R01 GM094800B (to G.J.J.), and HHMI (to M.K.W. and G.J.J.).

## References

- Alyahya SA, Alexander R, Costa T, Henriques AO, Emonet T, Jacobs-Wagner C. 2009. RodZ, a component of the bacterial core morphogenic apparatus. *Proc Natl Acad Sci* **106**: 1239–1244.
- Atmakuri K, Cascales E, Burton OT, Banta LM, Christie PJ. 2007. Agrobacterium ParA/MinD-like VirC1 spatially coordinates early conjugative DNA transfer reactions. *EMBO J* **26**: 2540–2551.
- Bendezú FO, Hale CA, Bernhardt TG, de Boer PA. 2009. RodZ (YfgA) is required for proper assembly of the MreB actin cytoskeleton and cell shape in *E. coli*. *EMBO J* **28**: 193–204.
- Ben-Yehuda S, Rudner DZ, Losick R. 2003. RacA, a bacterial protein that anchors chromosomes to the cell poles. *Science* **299**: 532–536.
- Bowman GR, Comolli LR, Zhu J, Eckart M, Koenig M, Downing KH, Moerner WE, Earnest T, Shapiro L. 2008. A polymeric



- protein anchors the chromosomal origin/ParB complex at a bacterial cell pole. *Cell* **134**: 945–955.
- Briegel A, Ortega DR, Tocheva EI, Wuichet K, Li Z, Chen S, Müller A, Iancu CV, Murphy GE, Dobro MJ, et al. 2009. Universal architecture of bacterial chemoreceptor arrays. *Proc Natl Acad Sci* **106**: 17181–17186.
- Cameron DE, Urbach JM, Mekalanos JJ. 2008. A defined transposon mutant library and its use in identifying motility genes in *Vibrio cholerae*. *Proc Natl Acad Sci* **105**: 8736–8741.
- Correa NE, Peng F, Klose KE. 2005. Roles of the regulatory proteins FlhF and FlhG in the *Vibrio cholerae* flagellar transcription hierarchy. *J Bacteriol* **187**: 6324–6332.
- Donnenberg MS, Kaper JB. 1991. Construction of an *ae* deletion mutant of enteropathogenic *Escherichia coli* by using a positive-selection suicide vector. *Infect Immun* **59**: 4310–4317.
- Ebersbach G, Ringgaard S, Møller-Jensen J, Wang Q, Sherratt DJ, Gerdes K. 2006. Regular cellular distribution of plasmids by oscillating and filament-forming ParA ATPase of plasmid pB171. *Mol Microbiol* **61**: 1428–1442.
- Ebersbach G, Briegel A, Jensen GJ, Jacobs-Wagner C. 2008. A self-associating protein critical for chromosome attachment, division, and polar organization in *Caulobacter*. *Cell* **134**: 956–968.
- Flärdh K. 2010. Cell polarity and the control of apical growth in *Streptomyces*. *Curr Opin Microbiol* **13**: 758–765.
- Fogel MA, Waldor MK. 2006. A dynamic, mitotic-like mechanism for bacterial chromosome segregation. *Genes Dev* **20**: 3269–3282.
- Heidelberg JF, Eisen JA, Nelson WC, Clayton RA, Gwinn ML, Dodson RJ, Haft DH, Hickey EK, Peterson JD, Umayam L, et al. 2000. DNA sequence of both chromosomes of the cholera pathogen *Vibrio cholerae*. *Nature* **406**: 477–483.
- Huitema E, Pritchard S, Matteson D, Radhakrishnan SK, Viollier PH. 2006. Bacterial birth scar proteins mark future flagellum assembly site. *Cell* **124**: 1025–1037.
- Kusumoto A, Shinohara A, Terashima H, Kojima S, Yakushi T, Homma M. 2008. Collaboration of FlhF and FlhG to regulate polar-flagella number and localization in *Vibrio alginolyticus*. *Microbiology* **154**: 1390–1399.
- Lam H, Schofield WB, Jacobs-Wagner C. 2006. A landmark protein essential for establishing and perpetuating the polarity of a bacterial cell. *Cell* **124**: 1011–1023.
- Lenarcic R, Halbedel S, Visser L, Shaw M, Wu LJ, Errington J, Marenduzzo D, Hamoen LW. 2009. Localisation of DivIVA by targeting to negatively curved membranes. *EMBO J* **28**: 2272–2282.
- Leonard TA, Butler PJ, Löwe J. 2005. Bacterial chromosome segregation: Structure and DNA binding of the Soj dimer—a conserved biological switch. *EMBO J* **24**: 270–282.
- Lutkenhaus J. 2007. Assembly dynamics of the bacterial MinCDE system and spatial regulation of the Z ring. *Annu Rev Biochem* **76**: 539–562.
- Lutkenhaus J. 2012. The ParA/MinD family puts things in their place. *Trends Microbiol* **20**: 411–418.
- Marston AL, Thomaidis HB, Edwards DH, Sharpe ME, Errington J. 1998. Polar localization of the MinD protein of *Bacillus subtilis* and its role in selection of the mid-cell division site. *Genes Dev* **12**: 3419–3430.
- Perez-Cheeks BA, Planet PJ, Sarkar IN, Clock SA, Xu Q, Figurski DH. 2012. The product of *tadZ*, a new member of the *parA/minD* superfamily, localizes to a pole in *Aggregatibacter actinomycetemcomitans*. *Mol Microbiol* **83**: 694–711.
- Radhakrishnan SK, Thanbichler M, Viollier PH. 2008. The dynamic interplay between a cell fate determinant and a lysozyme homolog drives the asymmetric division cycle of *Caulobacter crescentus*. *Genes Dev* **22**: 212–225.
- Ramamurthi KS, Losick R. 2009. Negative membrane curvature as a cue for subcellular localization of a bacterial protein. *Proc Natl Acad Sci* **106**: 13541–13545.
- Raskin DM, de Boer PA. 1997. The MinE ring: An FtsZ-independent cell structure required for selection of the correct division site in *E. coli*. *Cell* **91**: 685–694.
- Raskin DM, de Boer PA. 1999. Rapid pole-to-pole oscillation of a protein required for directing division to the middle of *Escherichia coli*. *Proc Natl Acad Sci* **96**: 4971–4976.
- Ringgaard S, Schirner K, Davis BM, Waldor MK. 2011. A family of ParA-like ATPases promotes cell pole maturation by facilitating polar localization of chemotaxis proteins. *Genes Dev* **25**: 1544–1555.
- Rudner DZ, Losick R. 2010. Protein subcellular localization in bacteria. *Cold Spring Harb Perspect Biol* **2**: a000307. doi: 10.1101/cshperspect.a000307.
- Schofield WB, Lim HC, Jacobs-Wagner C. 2010. Cell cycle coordination and regulation of bacterial chromosome segregation dynamics by polarly localized proteins. *EMBO J* **29**: 3068–3081.
- Semmler AB, Whitchurch CB, Leech AJ, Mattick JS. 2000. Identification of a novel gene, *fimV*, involved in twitching motility in *Pseudomonas aeruginosa*. *Microbiology* **146**: 1321–1332.
- Shapiro L, McAdams HH, Losick R. 2009. Why and how bacteria localize proteins. *Science* **326**: 1225–1228.
- Shiomi D, Sakai M, Niki H. 2008. Determination of bacterial rod shape by a novel cytoskeletal membrane protein. *EMBO J* **27**: 3081–3091.
- Szardenings F, Guymer D, Gerdes K. 2011. ParA ATPases can move and position DNA and subcellular structures. *Curr Opin Microbiol* **14**: 712–718.
- Takacs CN, Poggio S, Charbon G, Pucheault M, Vollmer W, Jacobs-Wagner C. 2010. MreB drives de novo rod morphogenesis in *Caulobacter crescentus* via remodeling of the cell wall. *J Bacteriol* **192**: 1671–1684.
- Thanbichler M. 2011. Good things come in small packages: Subcellular organization and development in bacteria. *Curr Opin Microbiol* **14**: 687–690.
- Thanbichler M, Shapiro L. 2006. MipZ, a spatial regulator coordinating chromosome segregation with cell division in *Caulobacter*. *Cell* **126**: 147–162.
- Viollier PH, Sternheim N, Shapiro L. 2002. A dynamically localized histidine kinase controls the asymmetric distribution of polar pili proteins. *EMBO J* **21**: 4420–4428.
- Wehbi H, Portillo E, Harvey H, Shimkoff AE, Scheurwater EM, Howell PL, Burrows LL. 2011. The peptidoglycan-binding protein FimV promotes assembly of the *Pseudomonas aeruginosa* type IV pilus secretin. *J Bacteriol* **193**: 540–550.
- Xu Q, Christen B, Chiu HJ, Jaroszewski L, Klock HE, Knuth MW, Miller MD, Elsliger MA, Deacon AM, Godzik A, et al. 2012. Structure of the pilus assembly protein TadZ from *Eubacterium rectale*: Implications for polar localization. *Mol Microbiol* **83**: 712–727.



## A multidomain hub anchors the chromosome segregation and chemotactic machinery to the bacterial pole

Yoshiharu Yamaichi, Raphael Bruckner, Simon Ringgaard, et al.

*Genes Dev.* 2012, **26**:

Access the most recent version at doi:[10.1101/gad.199869.112](https://doi.org/10.1101/gad.199869.112)

---

### Supplemental Material

<http://genesdev.cshlp.org/content/suppl/2012/10/11/26.20.2348.DC1>

### References

This article cites 40 articles, 17 of which can be accessed free at:  
<http://genesdev.cshlp.org/content/26/20/2348.full.html#ref-list-1>

### License

### Email Alerting Service

Receive free email alerts when new articles cite this article - sign up in the box at the top right corner of the article or [click here](#).

---

